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INTERNA	TIONAL APPLICATION NO. PCT/IT98/00231	INTERNATIONAL FILING DATE 11 August 1998	PRIORITY DATE CLAIMED 28 August 1997
TITLE C	E INIVENITION	MALS FOR THE STUDY OF BIOLOGICAL, PHYS	
	114110033110 1111	ria Grazia; ZECCA, Luigi; BROMLEY, Pete	
AFFLICA	CLERICI,	Libero A.; and VEZZONI, Paolo	
Applicant 1.	This is a FIRST submission of item. This is a SECOND or SUBSEQUE This express request to begin nation examination until the expiration of A proper Demand for International App a. X is transmitted herewith b. has been transmitted by c. is not required, as the a A translation of the International Amendments to the claims of the a. are transmitted herewith b. have been transmitted by c. have been transmitted by have been made; herewith have not been made; herewith have not been made; herewith have been made; herewith have been made; herewith have been made; herewith have not been made; herewith have been made; herewith herewith have been made; herewith have been made; herewith have been made; herewith herewith herewith have been made; herewith herewith have been made; herewith herewi	as Designated/Elected Office (DO/EO/US) the follows concerning a filing under 35 U.S.C. 371. NT submission of items concerning a filing under all examination procedures (35 U.S.C. 371(f)) at an examination procedures (35 U.S.C. 371(f)) at an examination procedure in 35 U.S.C. 371(f) at an explicable time limit set in 35 U.S.C. 371(f) at an explication as filed (35 U.S.C. 371(c)(2)) (Publication as filed (35 U.S.C. 371(c)(2)) (Publication was filed in the United States Receil Application was filed in the United States Receil Application into English (35 U.S.C. 371(c)(f) are International Application under PCT Article in (required only if not transmitted by the International Bureau. Designated/Elected Office (DO/EO/US) the International Bureau. Designated Office (DO/EO/US) the following such amend the will not be made.	35 U.S.C. 371. ny time rather than delay nd PCT Articles 22 and 39(1). onth from the earliest claimed priority date. No. WO 99/11772) national Bureau). eiving Office (RO/US). 2)). e 19 (35 U.S.C. 371(c)(3)) rnational Bureau).
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11. X	An Information Disclosure State	ement under 37 CFR 1.97 and 1.98.	
12.	An assignment document for rec	cording. A separate cover sheet in compliance	with 37 CFR 3.28 and 3.31 is included.
13. X	A FIRST preliminary amendmen	at.	
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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:)) Atty. Docket: SCBREV-223
SACCO, Maria Grazia, et al)
Serial No. (based on PCT/IT98/00231 filed on 11 August 1998)))))
Filed: Herewith	
For: TRANSGENIC ANIMALS FOR THE STUDY OF BIOLOGICAL, PHYSICAL AND CHEMICAL TOXIC AGENTS))) Date: 28 February 2000)

PRELIMINARY AMENDMENT

BOX: PCT (DO/EO/US)

Assistant Commissioner for Patents Washington, D. C. 20231

Sir:

Prior to calculating the filing fee, please amend the above-identified application as follows:

IN THE CLAIMS:

In the claims as amended during prosecution of corresponding international application, i.e. PCT/IT98/00231 (copy attached), please amend the claims as follows:

Claim 5, line 3, change "claims 1-4" to --claim 1--.

Claims 7, 8 and 9, line 1 of each, change "claims 5-6" to --claim 5--.

REMARKS

With the above amendments, the multidependencies of claims 5 and 7-9 have been deleted.

> Respectfully submitted, GRIFFIN, BUTLER, WHISENHUNT & SZIPL, LLP

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CLAIMS

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- 1. A non-human transgenic mammal which comprises cells containing a construct of a heat shock protein (hsp) promoter linked to the growth hormone (GH) gene sequence.
- A non-human transgenic mammal according to claim 1,
 wherein the heat shock protein promoter is hsp70 gene promoter.
 - 3. A non-human transgenic mammal according to claim 1, which is a rodent.
- A non-human transgenic mammal according to claim 3,
 which is a mouse.
 - 5. A method for the study of chemical, physical and biological toxic agents which comprises:
 - a) exposing the transgenic mammal of claims 1-4 to the toxic agent;
- 20 b) determining the effect through measurement of the hematic concentration of the reportergene.
 - 6. A method according to claim 5, wherein the same animal is used for repeated tests with the same or different toxic agent.
 - 7. A method according to claims 5-6, for the study of toxicity kinetics of one or more toxic agents.
 - 8. A method according to claims 5-6, for the study of heat stress.
- 30 9. A method according to claims 5-6, for the study of metal toxicity.

- 10. A method according to claim 9 for the study of toxicity of metals selected from the group consisting of Rb, Cu, Hg, As and Cd.
- 11. The use of the transgenic mammal of claim 1 for in vivo toxicity studies.

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12. The use of a transgenic animal according to claim 11, wherein said animal is a mouse.

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TRANSGENIC ANIMALS FOR THE STUDY OF BIOLOGICAL, PHYSICAL AND CHEMICAL TOXIC AGENTS

The present invention provides transgenic animals for the study of biological, physical and chemical toxic agents.

At present, toxicity tests can be carried out both in vivo and in vitro.

The industrials, the public opinion and the scientific community are strongly interested in the abolition of toxicity tests made on animals and therefore in their replacement with in vitro tests.

This target, however, is quite unrealistic at the moment, since no in vitro tests which can replace in vivo tests are available, either now or in the near future.

It is well known, in fact, that the substances under in vivo investigation often undergo metabolic modifications, which might significantly alter their toxicity profile, to an extent which would be unpredictable in in vitro tests.

On the other hand, in vivo studies always involve animal suffering and sacrifice.

However, it is possible to conceive genetically-engineered animal models which may simplify the determination of the toxicity of various agents and reduce the number of animals involved.

25 Recently, the use of transgenic animals as models for pharmacological studies has been proposed.

For example, EP 0 169 672 B1 describes transgenic animals bearing oncogenes like c-myc, suitable for the

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study of tumors associated to the expression of such oncogenes, or bearing the human growth hormone gene fused to a metallothionein promoter, whereby, said promoter being an inducible promoter, it is possible to study the effect of the expression, upon induction, of the associated gene on the whole organism (Palmiter et al. (1983) Science 222, 809).

WO 91/15579 describes a method for studying mutagenesis in transgenic animals bearing DNA sequences which can easily be extracted and analysed for mutations.

The present invention provides non-human transgenic animals useful for toxicity studies.

Such animals are characterised in that they have regulatory DNA sequences in some or all their cells, which are sensitive to biological, physical and chemical toxic agents, functionally linked to sequences of reporter genes, whereby the expression of the latter sequences is controlled or induced by said regulatory sequences.

Among the regulatory sequences, the stress-promoter sequences, like the heat shock protein (hsp) promoters, are preferred, but also cytochrome-promoters of the p450-superfamily, as well as those promoters of other genes, like p53 gene, activated by biological, chemical or physical stress, can be cited.

Among suitable reporter genes, the growth hormone gene, which has been used in the experiments described below, is preferred, but also chloramphenical acetyl transferase (CAT), green fluorescence protein (GFP) and β -galactosidase (LacZ) genes can be suitably employed.

The transgenic animals of the invention can be used

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in a method for studying the toxicity induced by various agents.

In theory, any animal normally suitable for a toxicity test can be used in the method of the invention. In practice, non-human mammals, particularly primates and rodents, are preferred.

Mice, in particular, are the most preferred.

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Conventional methods can be used for the production of transgenic animals, including, for example, the microinjection of recombinant DNA into embryonal cells or into pronuclei of one-cell stage embryos, the zygote, embryo cell, somatic cell or animal tissue infection with a virus, in particular with a retrovirus, according to what described, for example, in Hogan et al., Cold Spring Harbor Laboratory Press, NY, 1986; Palmiter et al., Ann. Rev. Genet., 20: 465-499; 1986; Capecchi, Science, 244: 288-292, 1989.

The method for the in vivo assay of potential toxic compounds according to the present invention, comprises exposing the animal to a chemical or physical agent for a time sufficient to induce the effect, and simply measuring the reporter gene expression. When the reporter gene encodes a protein secreted in the bloodstream, for instance, its hematic concentration, as well as other chemical-clinical parameters associated with the effect caused by the activation of the stress promoter, could be detected.

According to the first aspect of the invention, a preferred embodiment is the production of transgenic mice in which a construct has been inserted, which comprises a hsp promoter fused to growth hormone (GH)

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gene (transgene), said promoter being described in Dreano et al. (Biotechnology u6:953, 1988 and Gene 49:1-8, 1986) and in Fishbach et al. (Cell Biol. Toxicol. 9:177-188, 1993). The latter publication reports that the exposure to toxic metals of a stable fibroblast line, engineered with a construct containing the growth hormone gene under the control of hsp promoter, causes the secretion of the reporter gene in the medium.

According to the preferred embodiment of the invention, the injury caused by the toxic agent is determined as the increase of GH plasma concentration versus the control.

This model has resulted particularly efficient and sensitive, especially in relation with toxic metals, but it can suitably be used also for other classes of chemical toxic compounds, like endocrine disruptors, as well as for other physical or chemical agents, like radiations and electromagnetic fields.

The main advantages offered by the invention are: the possibility to diminish animal suffering, since only low amounts of the test substances are used, surely lower than the dosages which could induce suffering or death; the reduction of the number of animals used in toxicological tests; the provision of a model that is absolutely reliable for what concerns the metabolic modifications, which the toxic agents undergo in the organism, the interactions of toxic compounds with various organs and their final effects on cells, including the chronic effects. This model particularly useful for test resterations and allows to monitor the agent's effect during

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treatments using always the same animal, thus eliminating the variability of the individual response. Further, several compounds can be studied using the same animal. Finally, such transgenic models can be used also for in vivo studies of toxicity kinetics of toxic compounds.

The second aspect of the invention concerns the possibility to obtain primary cultures of cells from different tissues of the transgenic animal, in which a recombinant DNA construct is integrated as described above, whereby a cell- or tissue-specific toxicity study can be carried out and the intracellular biochemical effects connected to toxicity can be evaluated under controlled conditions and in more detail during different stages of animal growth.

In this case, the in vitro assay comprises preparing primary cultures in conditions variable depending on the cell type, exposing said cultures to the toxic agent and monitoring the activation of the stress promoter through detection of the protein encoded by the reporter gene.

Referring to the above described transgenic mice bearing the hsp/GH construct, an embodiment of the second aspect of the invention consists for example in preparing primary cultures of fibroblasts, kidney, lung or bone marrow cells, hepatocytes or other, in their simultaneous or separate treatment with one or more toxic agents, and in the determination of GH secretion in the medium.

If, using the above assay, a tissue or a cell-type resulted sensitive to the toxic agent, a deeper

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biochemical analysis could be made in order to find which cellular pathways are particularly involved in the toxicity.

Thus, according to a further aspect, the invention provides a method to carry out in vitro toxicity tests on primary cultures of somatic cells derived from a transgenic animal.

BRIEF DECRIPTION OF THE FIGURES

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Fig 1. Panel A: Southern blot analysis of transgenic heterozygous (lanes 1-4) and homozygous mice (lanes 5-7) and a non-transgenic control mouse (lane 8).

Panel B: RT-PCR with hGH specific primers of heat-shock activated liver cells from transgenic mice. Samples: RNA from cultured hepatocytes before (lane 1) and 30 min after (lane 2) heat shock in vitro; RNA from livers before (lane 3) and 30, 60, 90, minutes after heat shock (lanes 4-6). + and - represent the negative and positive controls respectively. Lanes 7 to 10 are the amplifications on non-retrotranscribed liver RNAs performed on the same samples as in lanes 3 to 6. M1: marker V, M2: 1 kb ladder.

Panel C: RT-PCR with HPRT specific primers performed on RNAs from the samples 1 to 6 as in panel B.

Fig. 2: Plasma levels of hGH (pg/ml) measured at different times in transgenic mice after thermal stress. Values represent the mean \pm SE; the number of mice tested for each time period is indicated by the number above each bar.

Fig. 3: Mean hGH plasma levels (pg/ml) ± SE observed in transgenic mice injected i.p. with PBS and with various inorganic toxic compounds at the indicated

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doses. Besides controls, are indicated: Rb: rubidium chloride; Hg: methylmercurium chloride; Cu: copper sulphate; Cd: cadmium chloride; As: sodium arsenite (2 doses)(below each bar is given the number of tested mice). The levels of significance are: *p<0.05; **p<0.01; ***p<0.005

Fig. 4: Mean ± SE of plasma hGH levels observed in transgenic mice subjected to two consecutive treatments, according to the following schema:

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Group	First treatment (T ₁)	Second treatment (T ₂)	Time Interval (T ₁ -T ₂)
As ₁	As	As	10 days
As ₂	Cđ	As	2 months
As ₃	Rb	As	2 months
Cu	Cu	Cu	2 months
Control	untretated	untreated	

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The following examples better illustrate the invention:

EXAMPLE 1

Production and characterization of a transgenic mouse lineage

25 <u>mouse lineage</u>

Transgenic mice were produced according to standard techniques (Hogan et al., "Manipulating the mouse embryo: a laboratory manual", Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1986), by microinjecting 1-cell stage embryo pronuclei with a 1.4 kb EcoRI DNA fragment from p17hGH construct (described in Dreano et al., Biotechnology 6:953, 1988 and Gene

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49:1-8, 1986), containing the human growth hormone cDNA as reporter gene, fused to the control region of the human Hsp70 promoter.

Mice were screened by Southern blot and/or PCR performed on tail DNA according to standard techniques. PCR was performed with the following primers: hGHL:GTGCAGTTCCTCAGGAGTGT; hGHR: CGAACTTGCTGTAGGTCTGC.

The amplification product was 171 bp long. Amplification conditions (35 cycles) were: 94°C for 20 sec, 58°C for 30 sec and 72°C for 20 sec. Heterozygous males and females were crossed and the homozygous progeny was identified by Southern blot, based on the intensity of the transgenic bands; their homozygosity was confirmed by checking the offspring when the homozygous male was mated to a non-transgenic partner. The mice used for the in vitro and in vivo experiments were always derived from a homozygous male bred with a non-transgenic CD-1 female.

Total RNA was extracted from different tissues

(liver, spleen, lung, kidney, blood) of transgenic and control mice, according to standard techniques (Sambrook et al., "Molecular cloning: a laboratory manual", Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.). Southern and Northern blot were performed according to standard techniques.

In order to evaluate the basal value of non-induced expression of the transgene, mice were analysed with Northern blot and with RT-PCR.

No expression was detected in lung, kidney, spleen,
liver and peripheral blood lymphocytes of non-treated animals or of animals not-exposed to heat shock. The hGH

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level in non-treated mice (control) was generally under the test detection limits, and when it was determined, it never exceeded 10 pg/ml.

EXAMPLE 2

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In vivo heat shock treatment.

Eight transgenic mice obtained according to example 1 and four non-transgenic control mice were subjected to in vivo heat shock at 44°C for 30 min. Six additional unexposed transgenic mice were tested. Aliquots of blood were taken before and 1, 3, 5, 7, and 24 hours after the heat shock.

In transgenic mice (Fig. 2) a specific increase of plasma hGH was detected with a peak three hour after treatment.

These results suggest that the integrated transgene does not affect in vivo the normal responsiveness of hsp promoter.

EXAMPLE 3

a) Inducibility of the hsp70/hGH transgene expression in vivo by sodium arsenite and methylmercurium chloride.

Male transgenic mice obtained as described in example 1 were weighed, anesthetized with ether and injected intraperitoneally (i.p.) with $NaAsO_2$ dissolved in PBS, at a final dose of 2.5 or 5 mg/kg, or with 3.5 mg/kg CH_3HgCl dissolved in PBS. Control transgenic mice were injected with the same volume of PBS (about 200 $\mu l/mouse$).

Blood samples were recovered before injection and 1, 3, 5, 7 and 24 hours after treatment.

30 hGH plasma levels at different times and doses are shown in Fig. 3.

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Both the tested doses of $NaAsO_2$ gave a clear and statistically significant response.

The response peaked after 3-5 hours and turned to the basal level 24 hours after injection.

- 5 CH₃HgCl gave hGH peaks after 5-7 hours and baseline hGH values 24 hours after injection.
 - b) Following the same procedure as described in a), hGH inducibility was evaluated in mice treated with rubidium chloride (18.5 mg/kg, c), copper sulfate (9 mg/kg, d) and cadmium chloride (4.7 mg/kg, e).

Results are reported in Fig. 3.

EXAMPLE 4

Inducibility of the hsp70/hGH transgene expression in vivo by repeated injections of toxic compounds.

15 Initially, 13 mice were treated as follows:

5 mice with As, 3 mice with Cd, 2 mice with Rb, 3 mice with Cu. After a period of 10 days to 2 months, the former three groups of mice were re-inoculated with As, the latter with Cu.

20 Blood samples were taken before and 3-5 hours after injection, i.e. at the times of highest response.

As shown in Fig. 4, after the first administration of the compound, the mice showed a response comparable to that observed in groups of mice treated as in example 3.

When retested after 10-60 days, a similar hGH increase was observed.

EXAMPLE 5

Embryonic fibroblast primary cultures-in vitro toxicity tests.

Homozygous transgenic mice obtained as described in

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example 1 were crossed with CD-1 females. After 14 days, embryonic fibroblasts (EMFIS) were recovered from the fetuses according to the technique described by Robertson E.J., IRL Press, Oxford, 77-88, 1987.

5 Cells were cultured in DMEM supplemented with 10% FCS and antibiotics (pen/strep), in an incubator (CO₂:5%, 100% humidity). Culture medium was replaced every second day with pre-warmed (37°C) fresh culture medium. The cells were expanded for two passages and then frozen at -80°C. For each experiment, cells were thawed, plated in 10 cm Petri dishes, left to grow and then re-seeded on 12 well plates until confluence.

To evaluate the toxic effect of the compounds, cells were treated by substituting the culture medium with fresh pre-warmed serum-free medium containing the toxic compounds at the chosen final dilutions. Cells were exposed to the toxic compound for either 5 or 24 hours and then the medium was replaced with fresh control medium for an additional 24 hours. At the end of the treatment, culture media were collected and assayed for hGH secretion by enzyme immunoassay.

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Each treatment was performed in triplicate and the hGH determination was repeated twice for each plate. The results are expressed as pg of hGH/ 10^6 cells. The sensitivity of this method was approximately 2-4 pg/ml.

As shown in the table, calcium and rubidium, known for their lack of toxicity at the tested concentrations, do not provoke hGH release in the medium.

On the contrary, a significant release is induced

30 after 24 hours of chrome exposure, while copper gives a
low response after 24 hours at the highest

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concentrations. On the contrary, mercurium does not induce hGH release from fibroblasts at each tested concentration.

Finally, arsenic and cadmium, as expected, showed clearly toxic.

EXAMPLE 6

Primary hepatocytes cultures-in vitro toxicity tests.

and their livers were perfused as described in Clerica et al., Mut. Res., 227:47-51, 1989, in order to collect hepatocytes. Hepatocytes were then seeded on 24 well plates (2x10⁵ cells/well) and cultured in William's E medium supplemented with antibiotics (pen/strep) and 10% FCS for 2 hours in order to allow them to attach to the bottom of the Petri dishes. The supernatant was then removed and the adherent cells were treated with the compounds dissolved in the medium.

To evaluate the toxic effect of the compounds, cells were treated by substituting the culture medium with fresh pre-warmed serum-free medium containing the toxic compounds at the chosen final dilutions.

As shown in the table, calcium and rubidium do not induce hGH release by mature hepatocytes.

25 Chrome treatment induces a high response after 24 hours, while copper treatment causes release either after 5 or 24 hours at each concentration.

Mercurium induces a response at concentrations higher than 5×10^{-5} M, while arsenic and cadmium show extremely toxic.

EXAMPLE 7

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In vitro toxicity tests on kidney, lung and bone-marrow primary cultures.

Kidney and lung cells were recovered as described by Campbell, J. A. et al. ("Sister cromatid exchange analysis of mice following in vitro exposure to vinyl carbonate", In vitro Cell. Dev. Biol. 22: 443:448, 1986).

Briefly, kidneys were removed from the same animals subjected to liver perfusion, washed 3 times in PBS additioned with antibiotics and minced in 0.5 mm pieces with a sterile scalpel. After 1 hour of incubation in trypsin/collagenase (100U/ml) solution, the suspension was centrifuged twice for 5 min. at 50xg, plated in 100 mm Falcon dishes and cultured in McCoy's medium with 20% FCS, 2mM Glutamine and Pen/strep.

In order to collect lung cells, after liver perfusion the chest cavity was opened after liver perfusion to access the lungs. The trachea was cut with a scalpel and a 22-gauge catheter was inserted into the trachea to perfuse the lungs with trypsin/collagenase solution for 5 min. in order to help the disaggregation of this tissue. The cells were then trypsinized, seeded in 24 wells and left to grow until confluence in McCoy's medium with 20% FCS, 2mM Glutamine and antibiotics.

In order to prepare bone marrow primary cultures, bone marrow cells were flushexd from the cavity of femurs and tibias with a syringe containing the culture medium. Cells were plated in 12 well plates with McCoy's medium with 20% FCS, 2mM Glutamine and antibiotics, and left to grow until the stromal cells reached confluence.

To evaluate the toxic effect of the compounds, the

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same procedure was followed as in the above examples 5 and 6.

Results are reported in the Table.

Table

(A) Determination of hGH (pg/10⁶ cells) release and primary transgenic cultures viability after 5-hour treatment

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Compounds	Primary lines	10 ⁻⁵ M	hGH rel 5x10 ⁻⁵ M	felease 5 _M 10-4 _M	5×10-4M	10-5M	Viabi 5x10 ⁻⁵ M	1ty 10-4M	5×10 ⁻⁴ M	
CaCl,	hepatocytes	nd	nd	pu	nd	+	+	+	+	
RbC1		nd	nd	nd	nd	+	+	+	+	
crcla	-	_	nd	nđ	nd	_	+	+	+	
CuSO4		_	pu	80	99		+	+	+	
K,Cr20,		nd	65	94	65	-/+	-/+	t	1	
сйінвсі		nđ	nd	nd	_	-/+	-/+	ı	_	
cdčl,		309	452	57	14	-/+	-/+	ı	. 1	15
$\mathtt{NaAs} \tilde{\mathtt{O}}_2$		100	224	nd	/	+	-/+	í	_	ĵ.
cacl,	Embryonic	_	nd	nd	nd	_	+	+	+	
RbC1	fibroblast	_	nd	nd	nd	_	+	+	+	
crcla		_	pu	nd	nd		+	+	+	
Cuso		_	nđ	9	12	/	+	+	+	
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cdčl,		250	85	45	nd	-/+	-/+	ı	. 1	
$\mathtt{NaAs\tilde{0}}_2$		pu	113	19	nd	+	-/+	-/+	/	

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CaCl,	Kidney		nd	nd	nd		-+	; + !	i i · +	ı
RbC1	cells	_	57	nđ	pu	_	+	+	+	
CrCla		_	pu	15	nd	_	+	+	-/+	
Cuso		_	nd	nđ	nd	_	+	-/+	-/+	
K2Cr207		nd	nd	pu	_	-/+	-/+	-/+	_	
сйынğсі		10	nd	nd	_	-/+	-/+	-/+	_	
cdč1,		nd	nd	nđ	_	-/+	ı	1	_	
$NaAs_2$		22	17	28	/	+	-/+	i	_	
CaCl ₂	Lungs	_	nd	127	202	_	+	+	+	
RbC1	cells	_	28	191	71	_	+	+	+	
CrClz		_	92	122	166	_	+	+	+	16
Cuso		_	nd	nd	184	_	-/+	-/+	-/+	
K2Cr207		pu	nd	nđ	_	-/+	1	1	_	
снічнёсі		27	nd	nd	_	i	1	ι	_	
cdč1,		nđ	31	11	/	-/+	ı	1	_	
$\mathtt{NaAs\tilde{o}}_2$		nd	37	249	/	-/+	-/+	1	/	

5-24-hour after measurable not were medium (controls) cells untreated in hGH levels

incubation.

nd = undetectable; / = not determined; + = with 100% viability; with 30-70% viability;

= 100% dead

17

(B) Determi	(B) Determination of hGH ((pg/10 ⁶	cells) r	release	and primary transgenic	trans		coltures a	ditei 64-
hour treatment.	ent.								
Compounds	Primary lines	10 ⁻⁵ M	hGH rel	lease 10-4M	5×10 ⁻⁴ M	10 ⁻⁵ M	Vital 5x10 ⁻⁵ M	ity 10-4 _M	5×10 ⁻⁴ M
CaC12 RbC1 CrC13 CuSO4 K2Cr207 CH3HGC1 CdC12	hepatocytes	nd nd / / nd nd nd	nd nd 36 12 nd 63 nd	nd nd 20 61 nd 103	nd nd 100 nd / 21	+ + \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	+ + + + 1 ! + +	++++1111	+ + 1 + 1 \ 1 \ -
CaCl2 Rbcl CrCl3 CuSO4 K2Cr2O7 CH3HGCl CdCl2	Embryonic fibroblast	/ / / nd / 181	nd nd 8 nd nd 108	nd nd 10 10 nd 41	ոգ ոժ ոժ ոժ դժ		++++11++	+ + + + 1 1 1 1	+ + + +

continues

	18		
1	10		24-hour
	1 1 + + + + 1 1 1 1	, ,	after
	+ + + + + 1 1 1 1		measurable
1 + +	1 1	1 1 1	not
			were
nd nd nd 450	nd 1114 35 901 /	nd 128 21 145 /	rols)
nd nd nd nd nd 110	nd 110 199 132 nd nd 415	51 20 21 127 127 165 nd	(cont
nd nd nd nd nd	nd 20 20 81 92 164 55	ոգ ոգ 38 ոգ ոգ	ls medium (controls)
/ / nd nd nd 300	/ / / / / / / / / / / / / / / / / / /	nd nd nd	cells
Kidney cells	Lungs cells	Bone marrow cells	els in untreated
Cacl2 Rbcl Crcl3 CuSO4 K2Cr207 CH3HGCl CdCl2	CaC12 RbC1 CrC13 CuSO4 K2Cr207 CH3H9C1 CGC12	CaCl2 Rbcl Crcl3 CuSO4 K2Cr2O7 CH3HGCl CdCl2	hGH levels

hGH levels in untreated cells medium (controls) were

CLAIMS

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- 1. A non-human transgenic mammal which comprises cells containing a construct of a heat shock protein (hsp) promoter linked to the growth hormone (GH) gene sequence.
- A non-human transgenic mammal according to claim 1,
 wherein the heat shock protein promoter is hsp70 gene
 promoter.
 - 3. A non-human transgenic mammal according to claim 1, which is a rodent.
- A non-human transgenic mammal according to claim 3,
 which is a mouse.
 - 5. A method for the study of chemical, physical and biological toxic agents which comprises:
 - a) exposing the transgenic mammal of claims 1-4 to the toxic agent;
- 20 b) determining the effect through measurement of the hematic concentration of the reporter-
 - 6. A method according to claim 5, wherein the same animal is used for repeated tests with the same or different toxic agent.
 - 7. A method according to claims 5-6, for the study of toxicity kinetics of one or more toxic agents.
 - 8. A method according to claims 5-6, for the study of heat stress.
- 30 9. A method according to claims 5-6, for the study of metal toxicity.

- 10. A method according to claim 9 for the study of toxicity of metals selected from the group consisting of Rb, Cu, Hg, As and Cd.
- 11. The use of the transgenic mammal of claim 1 for in vivo toxicity studies.
- 12. The use of a transgenic animal according to claim 11, wherein said animal is a mouse.







PCT

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Advanced Biomedical Technologies, Consiglio Nazionale delle Ricerche, Via Ampere, 56, I-20131 Milano (IT).

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(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

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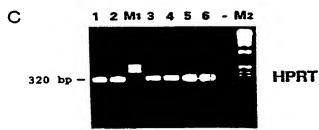
(54) Title: TRANSGENIC ANIMALS FOR THE STUDY OF BIOLOGICAL, PHYSICAL AND CHEMICAL TOXIC AGENTS

(57) Abstract

The invention provides non-human transgenic animals bearing regulatory DNA sequences in some or all their cells, which are sensitive to biological, physical and chemical toxic agents. Such sequences are linked to sequences of reporter genes useful for toxicological studies.

A 1 2 3 4 5 6 7 8

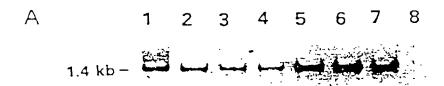
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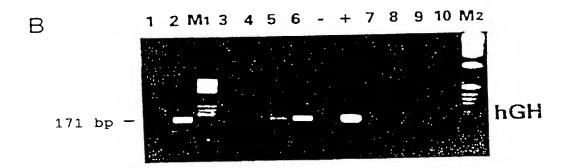


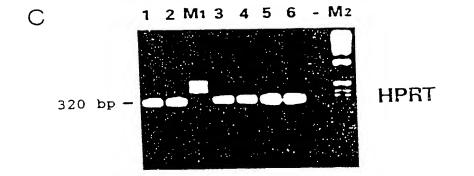
1/4

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FIGURE 1







2/4

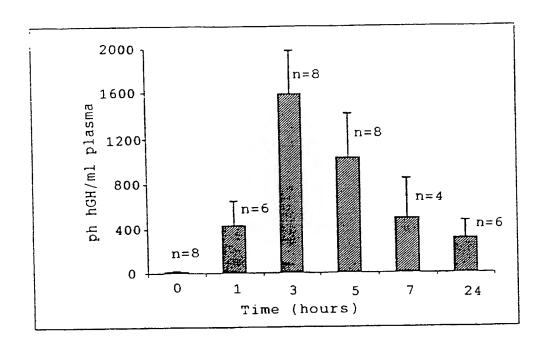
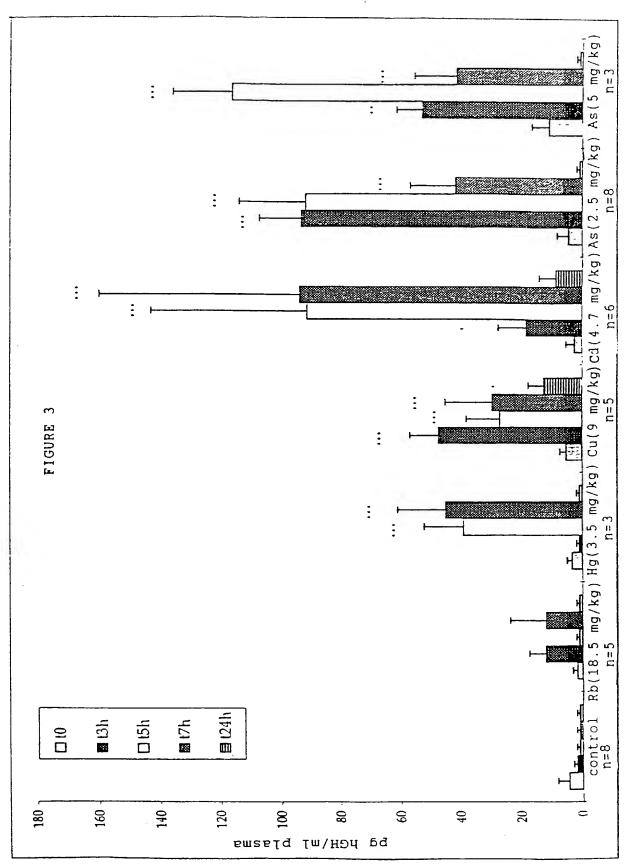


FIGURE 2

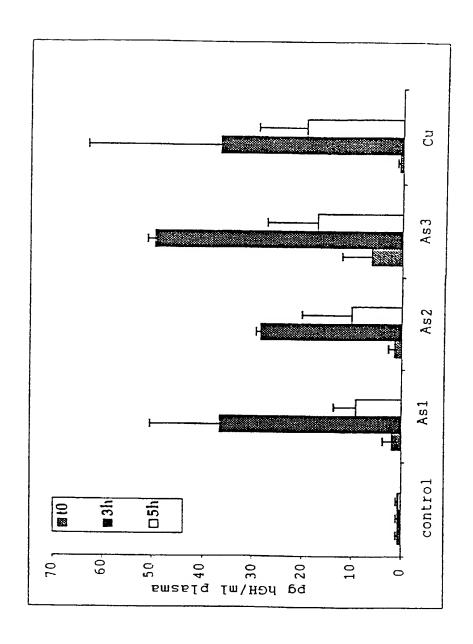
3/4



PCT/IT98/00231

4/4

FIGURE 4



(For Use With So	DECLARATION FOR UNITED S th PCT and Non-PCT Applicat	TATES PATENT APPLICATI Lions) (Atty. Docket:	ON SCBREV-223	<u></u>)
stated below next to my name, shelow) or an original, first is claimed and for which a pa or BICLOGUAL, PHYSICAL AND of which is attached hereto up	and joint inventor (if plu tent is sought on the inve HEMICAL TOXIC AGENTS	mal, first and sole in rel names are listed intion entitled, (1) 1 checked: (2) [xx] was	ventor (if only below) of the sul	one name is listed bject matter which s FOR THE STUDY the specification
I state that I have revi the claims, as amended by an which is material to patentab 119 of any foreign application below any foreign application application on which priority	ility as defined in 37 CFF m(s) for patent or inventor n for patent or inventor	bove. I acknowledge R 1.56 I claim forai or's certificate list	the duty to dis ign priority bene ed below and hav	sclose information efits under 35 USC we also identified
Prior Foreign Application(s)	* •			
NUMBER	COUNTRY	DAY/MONTH/YEAR FI	T FIRE	DIAMETER OF STREET
(6) <u>MI97A (101972</u>	TTALY	28 August 1997		RIORITY CLAIMED [X] Yes [] No
PCT/IT98/00231	PCT	11 August 1998		(X) Yes [] No
				[] Yes [] No
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subject matter of each of application in the manner pro- information which is material filing date of the prior appli-	wided by the first paragr to patentability as defi	ication is not disclorable of 35 USC 112, I ned in 37 CFR 1 56 W	osed in the pri acknowledge the	ior United States duty to disclose
Appln. Sorial No.	Filing Date	• •		
(7)		(Status: [] Patente (Status: [] Patente	ed [] Pending [Abandoned)
I appoint B. Franklin	Griffin, Jr., Reg. No.			
I appoint B. Franklin Whisenburt, Reg. No. 24,378; individually and jointly my application and to transact a resulting patent, whose addr. South, Arlington, Virginia 22.				
	ements made herein of my of lieved to be true; and fur and the like so made are	own knowledge are true Ther that these states	and that all a	statements made on with the knowledge
(8) Full name of sole or first	inventor SACO Maria	Gradia	•	
Inventor's signature	a hour to	GIBZIG	Date 21 1	arch toco
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56, 20131 Milano. Italy	- }	Citizens	ship <u>Italian</u>	
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56, 20131 Milano, Italy	TX		shipItalian	
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(1) Insert title of invention (2) Check block for PCT appli If FCT national phase entry of	cation or U.S. application oplication of U.S. application, insert internal	on already on file, ar cional PCT application	nd complete items filing date, Se	3 (3),(4) and (5).
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(7) Complete for earlier US pa (8) Complete ALL blanks, Attac	rent applications; and add	litional page if needed	i.	
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(8) Full name of joint invantor CLERICI, Libero	Date 23 Marko 200
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Inventor's signature	TT Citizenship Italian
	LIA CICIZENDITY
Post Office Address Same as Above	
(8) Full name of joint inventor VEZZONI, Faolo	2000 Land Flora
	Date 17 (Steelers)
Inventor's signature 11200 Technologies Consignation Institute of Advanced Biomedical Technologies Consignation	<u>io Nazionala delle Ricerche. Via Amper</u>
Residence Institute of Mayantan	Citizenship Italian ———
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Inventor's signature	
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(8) Full name of joint inventor BROTALY FETCH	Date 31 Mort
Pater Drawbay	
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CH-1211, GENEVA 24, SWITZER LAND	Citizenship DRITISH
POSC VILICE Address	
(8) Full name of joint inventor	Date
Inventor's signature	
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Post Office Address	
(8) Full name of joint inventor	Date
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(8) Full name of joint inventor	Date
Inventor's signature	
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Post Office Address	
(8) Full name of joint inventor	
	pate
Inventor's signature	
Residence	Citizenship

·	DECLARATION FOR UNITED STATES PATENT APPLICATION (For Use with Both PCT and Non-PCT Applications) (Atty Docket: SCHREY-223)
	As the below named legal guardian of Rachele RONCUCCI and Ragine RONCUCCI, I declare that Rachele RONCUCC
· · ·	and Regime RONCUCCI are minor heirs of Romeo RONCUCCI, deceased, who is an inventor/applicant of the subject
	matter which is claimed and for which a patent is sought on the invention entitled, (1) <u>TRANSGENIC ANIMALS R</u> THE STUDY OF BIOLOGICAL PRYSICAL AND CHEMICAL TOXIC AGENTS
1	specification of which is attached hareto unless the following box is checked: (2) [XX] was filed on (3) = February 2000 (4) as U. S. Appl. SN or FCT International Appl. No.
1 2	and was amended on (5) 28 February 2000 (1f applicable).
eta de	I state that I have reviewed and understand the contents of the above-identified specification
	including the claims, as amended by any amendment referred to above. The duty to disclose information which is material to patentability as defined in 37 CFR 1.56 is hereby acknowledged. Foreign priority benefit under 35 USC 119 of any foreign amplication(s) for patent or inventor's certificate listed below are heart
*	claimed and identified below are any foreign application for patent or inventor's certificate having a filir date before that of the application on which priority is claimed:
	Prior Foreign Application(s)
•	(6) MI97A 001972 ITALY 28 August 1997 [X] Yes [I No
t	PCT 11 August 1998 [X] Yes [] No
	[] Yes [] No
	I claim the benefit under 35 USC 120 of the United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United State application in the manner provided by the first paragraph of 35 USC 112, I acknowledge the duty to disclose
-3	
2.5	filing date of the prior application and the national or PCT international filing date of this application:
	Applm. Serial No. Filing Date
7	(Status: [] Patented [] Pending [] Abandoned)
	(Status: [] Patented [] Pending [] Abandoned)
	I appoint B. Franklin Griffin, Jr., Reg. No. 19,334; F. Prince Butlor, Reg. No. 25,566; Fred S. Whisenbunt, Reg. No. 24,378; Joerg-Uwe Szipl, Reg. No. 31,799; and Richard J. Gallagher, Reg. No. 28,761
	resulting patent, whose address is Griffin, Butler, Whisenhunt & Szipl, Lip, Suite PH-1, 2300 9th Street South, Arlington, Virginia 22204-2320, Telephone No. (703) 979-5700, Customer No. 113.
100	I declare that all statements made herein of my own broadedge and have and that
	information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under I
	ose tool and that such willful raise statements may leopardize the validity of the application or any paten
	issued thereon.
	(n) Romeo RONCUCCI, Decessed
	Ψ°
100	By Marie Moulle Carl
53.55	Maria Novella CASTAGNOLL, Guardian of Rachele
	RONCUCCI and Regime RONCHCCT who are miner
	heirs of Romeo RONCUCCI, Decessed, and who reside at via Ungaretti 17, 20028 San Vittore Olona, Milang Iraly
1	Milano, Italy
	TTX
·	Dated: 23.05
	(1) Insert title of invention. (2) Check block for PCT application or U. S. application already on file, and complete items (3),(4) and (5). If PCT national phase matrix application is application already on file, and complete items (3),(4) and (5).
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DECLARATION FOR UNITED STATES PATENT APPLICATION  (For Use With Both PCT and Non-PCT Applications) (Atty. Docket: <u>SCEREY-323</u> )
I declare that I am the below named heir of Romeo RONCUCCI, deceased, who is an inventor/applicant of the subject matter which is claimed and for which a patent is sought on the invention entitled, (1) TRANSCENIC ANIMALS FOR THE STUDY OF BIOLOGICAL PHYSICAL AND CHEMICAL TOXIC AGENTS  the specification of which is attached hereto unless the following box is checked: (2) [XX] was filed on (3) 28  February 2000 (4) as U. S. Appl. SN or FCT International Appl. No.
and was amended on (5) 28 February 2000 (if applicable).  I state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above. The duty to disclose information which is material to patentability as defined in 37 CFR 1.56 is hereby acknowledged. Foreign priority benefits under 35 USC 119 of any foreign application(a) for patent or inventor's certificate listed below are hereby claimed and identified below are any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed.
Prior Foreign Application(s)    MIMBER   COUNTRY   DAY/MONTH/YEAR FILED   PRIORITY CLAIMED
I claim the benefit under 35 USC 120 of the United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of 35 USC 112, I acknowledge the duty to disclose application which is material to patentability as defined in 37 CFR 1.56 which became available between the filling date of the prior application and the national or PCT international filling date of this application:
Applu, Serial No. Filing Date  (Status: [ ] Patented [ ] Pending [ ] Abandoned)  (Status: [ ] Patented [ ] Pending [ ] Abandoned)
I appoint B. Franklin Griffin, Jr., Reg. No. 19,334; F. Princo Butler, Reg. No. 25,666; Fred S. Whisenhunt, Reg. No. 24,378; Joseg-Uwe Szipl, Reg. No. 31,799; and Richard J. Gallagher, Reg. No. 28,781, individually and jointly my attorneys with full power of substitution and revocation, to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith, and with the resulting patent, whose address is Griffin, Butler, Whisenhunt & Szipl, LLP, Suite PH-1, 2300 9th Street, South, Arlington, Virginia 22204-2320, Telephone No. (703) 979-5700, Customer No. 113.
I declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under 18 that willful false statements and the like so made are punishable by fine or imprisonment or both, under 18 to 1001 and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.
Romeo RONCUCCI, Decoased
Sylvie KONCUCCI ar heir of Romco RONCUCCI, Deceased, residing at Via Thaon di Revel, 12, 20159 Milano, Italy
Dated: 23.05 , 2000
<ol> <li>Insert title of invention.</li> <li>Check block for PCT application or U. S. application already on file, and complete items (3), (4) and (5).</li> <li>Check block for PCT application or U. S. application already on file, and complete items (3), (4) and (5).</li> <li>CCT national phase entry application, insert international PCT application filing date, Serial No., and date of any Article 19 amendments.</li> <li>Complete for foreign priority documents; add additional page if needed.</li> <li>Complete for earlier US parent applications; add additional page if needed.</li> </ol>

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